Background

Surgical site infection (SSI) are common complications of surgical care and account for 14% to 16% of all nosocomial infections. They have a high impact on morbidity, mortality in surgical and intensive care units (ICU). In order to establish the prophylaxis protocols, we must know the antibiotic resistance profiles of the germs involved in SSI.

Objectives

1. Establishing the etiology of SSI in post-surgery patients.
2. Comparison of antibiotic resistance profile variation in relation to the suppuration and ward type (surgical or ICU).

Material and method

Study lot: 165 hospitalized patients from Clinical Emergency County Hospital of Craiova, Romania between 1.06-31.08.2014.

Biological samples: discharge, pus, drainage liquid from superficial and deep surgical wounds.

We performed classical bacteriological diagnosis and antimicrobial susceptibility testing by diffusimetric method (Clinical and Laboratory Standards Institute's guidelines 2014).

The results were processed using Whonet 5.6 software. Statistical analyses were performed using SPSS 20.0.

Antimicrobial susceptibility testing by diffusimetric method

Results

- **Bacterial species involved in surgical site infections**
- **Variation of multiple antibiotic resistance index (MAR) by species and type of ward**
- **Antibiotic resistance profile of Staphylococcus aureus**
- **Antibiotic resistance profile of Klebsiella pneumoniae**
- **Antibiotic resistance profile of Staphylococcus aureus in surgery and ICU wards**
- **Antibiotic resistance profile of Klebsiella pneumoniae in surgery and ICU wards**
- **Antibiotic resistance profile of Escherichia coli**
- **Hierarchical cluster analysis of resistance of Escherichia coli and Klebsiella pneumoniae strains (Ward’s minimum variance method)**
- **Antibiotic resistance profile of Escherichia coli in surgery and ICU wards**
- **Hierarchical cluster analysis of resistance of Staphylococcus aureus strains (Ward’s minimum variance method)**

Discussion

The clinical hospital pharmacist play an important role in the antibiotic stewardship of SSI by monitoring infection rates, participating in the selection of antibiotics, and determining when to initiate, de-escalation and terminate antibiotic prophylactic administration. Clinical hospital pharmacist must ensure that the number and types of antimicrobial agents available are appropriate for the patient population served (wards, geographic area, disease type).

The drug sensitivity profile (diameters of inhibition areas) in the diffusimetric method can be used for phenotypic typing and epidemiological tracing of hospital strains.

Conclusions

The study identified the antimicrobial resistance profiles of main bacterial agents involved in SSI.

The germs isolated from ICU wards have a higher resistance to certain antimicrobials, especially to the carbapenems, compared with those from surgical wards. The use of these agents should be restricted in ICU wards to prevent the development of the resistance.

The clinical hospital pharmacist must work closely with the surgeons, epidemiologists, infectionists and microbiologists for elaboration of clinical infection prevention guidelines.